



UNIVERSITI PUTRA MALAYSIA

***BIODEGRADATION OF DIESEL FUEL BY TWO PSYCHROTOLERANT
STRAINS ISOLATED FROM SOUTHERN VICTORIA ISLAND,
ANTARCTICA***

NUR MUHAMAD SYAHIR BIN ABDUL HABIB

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ANTARCTICA**

NUR MUHAMAD SYAHIR BIN ABDUL HABIB

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirement for the Degree of Master of Science**

November 2017

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DEDICATION

This thesis is dedicated to my parents.



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Abstract of thesis presented to Senate of Universiti Putra Malaysia in fulfillment
of the requirements for the degree of Master of Science

**BIODEGRADATION OF DIESEL FUEL BY TWO PSYCHROTOLERANT
STRAINS ISOLATED FROM SOUTHERN VICTORIA ISLAND,
ANTARCTICA**

By

NUR MUHAMAD SYAHIR BIN ABDUL HABIB

November 2017

Chairman: Nur Adeela Binti Yasid, PhD

Faculty: Biotechnology and Biomolecular Sciences

Hydrocarbon contamination in Antarctica poses a great threat to the delicate and unique ecosystems of this continent. Bioremediation of hydrocarbon pollutants via utilisation of the indigenous hydrocarbon-degrading bacteria, has been proposed as an environmentally friendly method to clean-up contaminated soils in Antarctica. This study focused on diesel-degrading *Pseudomonas* and *Rhodococcus* species isolated from pristine soils located at the Southern Victoria Island, Antarctica. Isolates were assessed for their ability to grow on diesel as a sole carbon source on solid media at 4°C. Nine isolates showed significant growth in enriched agar after 14 days of incubation. Isolates were then screened to obtain the most promising diesel-degrading strains through colourimetric assay. Two potent isolates that possess rapid utilisation of 0.5% (v/v) diesel were selected and identified as *Pseudomonas* sp. strain ADL15 and *Rhodococcus* sp. strain ADL36. The factors that contribute to the growth of both strains were characterised initially using the conventional ‘one-factor-at-a-time’ approach. During this stage, the optimal condition for the growth of both ADL15 and ADL36 were at pH 7.0, 20°C, 1.0% (w/v) NaCl, and 1.0 g/L NH₄NO₃. However, strain ADL36 favoured a higher amount of diesel (2.0% (v/v)) for bacterial growth by comparison to ADL15 (1.0% (v/v)). Percentage of dodecane mineralisation was used as the mean to indicate diesel reduction through gas chromatographic analysis. While strain ADL36 showed 83.75% of dodecane mineralisation, the reduction of dodecane by AD15 is merely at 22.39%. Response surface methodology (RSM) based on central composite design (CCD) was used to improve and optimise the effect of significant factors toward the biodegradation of diesel. RSM proved to enhance the reduction of experimented hydrocarbon (dodecane) with a 15% and 16% increment of mineralisation for isolate ADL15 (38.32%) and ADL36 (99.89%), respectively. The results also demonstrated that addition of salt to culture media was the limiting factor in hydrocarbon degradation. Whole genome sequencing showed that ADL15 and ADL36 were closely related to the *Pseudomonas fluorescens* and *Rhodococcus erythropolis* grouping, respectively. Metagenomic analyses revealed the presence of alkane hydroxylases systems which was responsible for alkane degradation in ADL36 but not in ADL15. This finding corresponds to the

gas chromatographic analysis in which ADL36 proved to be a better alkane degrader than ADL15. Detection of the complete pathway of aromatic compound degradation in the latter strain indicates a stronger inclination of the strain to utilise aromatic components in diesel as the carbon source. The presence of putative monooxygenases may also suggest that this strain may utilise specific alkane for their growth. The results from this study showed that strain ADL15 and ADL36 have an excellent potential in bioremediation of aromatics and aliphatics, respectively.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

**BIODEGRADASI MINYAK DIESEL OLEH DUA STRAIN PSIKROTOLERAN
YANG DIPENCILKAN DARIPADA PULAU VICTORIA SELATAN,
ANTARTIKA**

Oleh

NUR MUHAMAD SYAHIR BIN ABDUL HABIB

November 2017

Pengerusi : Nur Adeela Binti Yasid, PhD
Fakulti : Bioteknologi dan Sains Biomolekul

Pencemaran hidrokarbon di Antartika menyebabkan ancaman yang besar kepada ekosistem yang unik dan rapuh di benua ini. Bioremediasi yang menggunakan bakteria yang mampu menguraikan hidrokarbon, telah dicadangkan sebagai kaedah yang mesra alam bagi membersihkan kawasan tercemar di Antartika. Kajian ini memberi tumpuan kepada pengurai minyak diesel daripada spesis *Pseudomonas* dan *Rhodococcus* yang diasinkan daripada tanah yang suci yang terletak di Pulau Victoria Selatan, Antartika. Pencilan dinilai melalui keupayaan mereka untuk menumbuh dengan menggunakan diesel sebagai sumber karbon tunggal pada media pepejal pada 4°C. Sembilan pencilan menunjukkan pertumbuhan yang jelas di dalam agar yang diperkaya selepas 14 hari inkubasi. Pencilan kemudian disaring untuk mendapatkan pencilan pengurai minyak diesel yang paling bagus melalui ujian berwarna. Dua pencilan kekar yang menggunakan 0.5% (v/v) minyak diesel secara pantas telah dipilih dan dikenalpasti sebagai *Pseudomonas* sp. strain ADL15 dan *Rhodococcus* sp. strain ADL36. Faktor-faktor yang menyumbang kepada pertumbuhan kedua-dua strain dicirikan pada mulanya menggunakan pendekatan konvensional ‘satu-faktor-pada-satu-masa’. Pada peringkat ini, keadaan optimum pertumbuhan ADL15 dan ADL36 berada pada pH 7.0, 20°C, 1.0% (w/v) NaCl, dan 1.0 g/L NH₄NO₃. Walau bagaimanapun, strain ADL36 menyukai jumlah minyak diesel yang lebih tinggi (2.0% (v/v)) bagi pertumbuhan bakteria berbanding ADL15 (1.0% (v/v)). Peratusan penguraian dodekana digunakan sebagai tanda bagi menunjukkan pengurangan diesel melalui analisis kromatografi gas. Walaupun strain ADL36 menunjukkan 83.75% penguraian dodekana, pengurangan dodekana oleh ADL15 adalah hanya pada 22.39%. Pengkaedahan tindakbalas permukaan (RSM) berdasarkan reka bentuk komposit pusat (CCD) digunakan untuk meningkatkan dan mengoptimumkan kesan faktor-faktor penting kearah penguraian diesel. RSM terbukti dapat meningkatkan pengurangan hidrokarbon (dodekana) yang diuji dengan peningkatan sebanyak 15% dan 16% untuk penurunan dodekana bagi pencilan ADL15 (38.32%) dan ADL36 (99.89%), masing-masing. Keputusan yang diperoleh juga menunjukkan bahawa penambahan garam ke media kultur adalah faktor yang mengurangkan penguraian hidrokarbon. Penujukan keseluruhan genom menunjukkan bahawa ADL15 dan ADL36 mempunyai kaitan rapat dengan kelompok *Pseudomonas fluorescens* dan *Rhodococcus erythropolis*. Analisis metagenomik mendedahkan

kehadiran sistem alkane hidroksilase yang bertanggungjawab terhadap penguraian alkana di dalam ADL36 tetapi tidak di dalam ADL15. Penemuan ini bersesuaian dengan analisis kromatografi gas dimana ADL36 terbukti menjadi pengurai alkana yang lebih baik daripada ADL15. Pengesahan laluan penguraian hidrokarbon aromatik yang lengkap didalam ADL15 mungkin menunjukkan kecenderungan ADL15 yang lebih kuat untuk menggunakan bahagian aromatik didalam diesel sebagai sumber karbon. Kehadiran monoooksigenase putatif mungkin juga menunjukkan bahawa strain ini menggunakan alkana yang khusus untuk pertumbuhan mereka. Keputusan daripada kajian ini menunjukkan bahawa strain ADL15 dan ADL36 mempunyai potensi yang baik didalam bioremediasi hidrokarbon aromatik dan alifatik.

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Syahir Habib, 2017

I certify that a Thesis Examination Committee has met on 30 November 2017 to conduct the final examination of Nur Muhamad Syahir bin Abdul Habib on his thesis entitled "Biodegradation of Diesel Fuel by Two Psychrotolerant Strains Isolated from Southern Victoria Island, Antarctica" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Syahida binti Ahmad, PhD

Senior Lecturer

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

Noor Azmi Shaharuddin, PhD

Senior Lecturer

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Internal Examiner)

Sazlina Md Salleh, PhD

Senior Lecturer

Universiti Sains Malaysia

Malaysia

(External Examiner)



NOR AINI AB. SHUKOR, PhD

Professor and Deputy Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 27 February 2018

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirements for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Nur Adeela Yasid, PhD

Senior Lecturer

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairperson)

Siti Aqlima Ahmad, PhD

Senior Lecturer

Faculty Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

Mohd Yunus Shukor, PhD

Associate Professor

Faculty of Biotechnology

Universiti Putra Malaysia

(Member)

Wan Lutfi Wan Johari, PhD

Senior Lecturer

Faculty of Environmental Sciences

Universiti Putra Malaysia

(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

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Signature: _____
Name of Chairman
of Supervisory
Committee: Dr. Nur Adeela Yasid

Signature: _____
Name of Member
of Supervisory
Committee: Dr. Siti Aqlima Ahmad

Signature: _____
Name of Member
of Supervisory
Committee: Associate Professor
Dr. Mohd Yunus Shukor

Signature: _____
Name of Member
of Supervisory
Committee: Dr. Wan Lutfi Wan Johari

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LIST OF ABBREVIATIONS

%	Percent
% (v/v)	Percent concentration volume / volume
% (w/v)	Percent concentration weight / volume
°C	Degree celsius
µl	Microlitre
µm	Micrometre
x g	Relative centrifugal force
bp	Base pair
CCD	Central composite design
CFU	Colony forming unit
DNA	Deoxyribonucleic acid
dNTP	Deoxyribonucleotide phosphate
EDTA	Ethylenediaminetetraacetic acid
EtBr	Ethidium bromide
et al.,	And friends
g	Gram
GC-FID	Gas chromatography- flame ionisation detector
gDNA	Genomic DNA
g/L	Gram per Litre
HCl	Hydrochloric acid
K	Kelvin
kb	Kilobase
L	Litre
M	Molar
min	Minute
mg/ml	Milligram per millilitre
mM	Milimolar
mm	Millimetre
ng/µl	Nanogram per microlitre
nM	Nanomolar
nm	Nanometer
OD	Optical density
OFAT	One-factor-at-a-time
PAHs	Polycyclic aromatic hydrocarbons
RNA	Ribonucleic acid
rpm	Revolution per minute
rRNA	Ribosomal RNA
RSM	Response surface methodology
sp.	Species (singular)
TAE	Tris-acetate-EDTA

CHAPTER 1

INTRODUCTION

Oil and its refined products represent a significant fraction of the pollution found in the Antarctic region, an area considered as the most pristine in the world (Raymond et al., 2017). The occurrences of pollution are more clustered near former military and industrial spots, scientific research stations, rural communities, and remote airfields, while recent spills and discharges tend to be linked with resource development and mishaps in transportation (Aislabie et al., 2004). Fuel spills are recognised as a potential threat as contamination can cause risks for humans and other living organisms if oil and fuel reach groundwater reservoirs and water bodies (Wang and Bartha, 1990; Jesus et al., 2015). Hydrocarbon contamination in these ecosystems is perceived as damaging as they are more sensitive, being profoundly adapted to extreme conditions (McDonald and Knox, 2014; Yang et al., 2009). Besides, due to the slow natural attenuation rates in cold climates, hydrocarbons can persist for longer periods of time than their temperate counterparts causing a stunted ecosystem recovery (Snape et al., 2008). Although the crude oil extraction from the polar region is declining and a strict legislation such as Antarctic Treaty was introduced, contamination of hydrocarbons in the Antarctic region may still be introduced by the booming numbers of tourists during the austral summer (November to March) period. According to the recent tourism statistics recorded by the International Association of Antarctica Tour Operators (IAATO), there is a 16% increase in the number of tourists landed on Antarctic from the 2015-2016 to 2016-2017 tour (IAATO 2016, 2017). While tourism cannot be solely blamed for their chances of introducing contamination, any small contamination can cause great risks to the environment for a remote and almost pristine land such as Antarctica.

Soil remediation in Antarctica is driven by several critical factors such as cost, strict environmental policy, and remediation constraints (Filler et al., 2006). Attempts to clean-up Antarctic polluted sites using both physical and chemical methods have been done but considered as a minor success. Several methods which highly practical in temperate environments such as thermal incineration, is banned from the Antarctic environment while soil excavation and removal of contaminated soils are often impractical, for the reason of high cost and risks of further damage from excavation (Snape et al., 2008). Bioremediation is widely proposed to remove pollutants from the contaminated Antarctic environment due to the increased interest in using the eco-friendly method as a process of remediating diesel fuel polluted sites (Aislabie et al., 2006; Jesus et al., 2015; Rayner et al., 2007). Bioremediation aids remediation activities being carried out either near or on site, which can be appealing in an isolated contamination spot. However, the effectiveness of this approach depends on strong limitations in temperature, bioavailability, oxygen, toxicity, and soil freeze-thaw cycle (Yang et al., 2009; Delille and Coulon, 2008). Among the factors, temperature plays a significant role in determining the rate and degree of microbial hydrocarbon biodegradation while affecting the volatilisation and viscosity of hydrocarbons (Delille and Coulon, 2008).

Biodegradation of varied components of hydrocarbons at low temperatures in Antarctic soils (Baraniecki et al., 2002; Bej et al., 2000) has been reported and is a result of the degradation capacity of indigenous cold-adapted microorganisms. Cold-adapted microorganisms are able to grow at temperatures around 0°C and have adapted their metabolism to function optimally at low temperatures. These microorganisms play a substantial role in the *in situ* biodegradation of hydrocarbons in cold environments, where ambient summer temperatures often correspond with their growth temperature range. As the Antarctic Treaty prohibits the introduction of non-native organisms, microbes that are indigenous to the Antarctic soil were required for the application of bioremediation (Aislabie et al., 2000). Among microbes, bacterial species play a key role in degrading hydrocarbon pollutants. A large number of hydrocarbon-degrading bacteria from cold soils have been identified, including representatives of gram-negative and gram-positive genera (Aislabie et al., 2000; Ruberto et al., 2005; Shukor et al., 2009). Although large numbers of hydrocarbon-degrading bacteria were isolated from contaminated soils, Margesin et al. (2003) and Stallwood et al. (2005) have observed the occurrence of bacterial species with hydrocarbon-degradative ability in the pristine soil. The addition of indigenous bacteria isolated from Antarctic pristine soil that possess a high competency to degrade diesel fuel may speed up the mineralisation process of petroleum hydrocarbons by several folds when favourable condition is maintained and the regulation of genetic diversity in the bacteria is acknowledged.

Thus, a study was carried out with the following objectives:

1. To isolate and identify bacterial species with hydrocarbon-degrading capacity from the Antarctic pristine soil.
2. To determine the optimum condition for bacterial growth and dodecane mineralisation for isolated strain using statistical analysis and model prediction.
3. To observe the residual hydrocarbon compounds in culture-optimised state qualitatively using gas chromatographic analysis.
4. To analyse the responsible alkane pathways, hydrocarbon-degrading enzymes and their respective genes through bacterial whole genome sequencing.

REFERENCES

- Abdel-Shafy, H. I., & Mansour, M. S. M. (2016). A review on polycyclic aromatic hydrocarbons: source, environmental impact, effect on human health and remediation. *Egyptian Journal of Petroleum*, 25(1), 107–123.
- Abd-Elsalam, K. A. (2003). Bioinformatic tools and guideline for PCR primer design. *African Journal of Biotechnology*, 2(5), 91–95.
- Abedon, S. T. (2012). Bacterial 'immunity' against bacteriophages. *Bacteriophage*, 2(1), 50–54.
- ACGIH (American Conference of Governmental Industrial Hygienists). (2005). Polycyclic aromatics hydrocarbons (PAHs) biologic exposure indices (BEI) Cincinnati.
- Aggarwal, R. K., Dawar, C., Phanindranath, R., Mutnuri, L., & Dayal, M. (2016). Draft genome sequence of a versatile hydrocarbon-degrading bacterium, *Rhodococcus pyridivorans* strain KG-16, collected from oil fields in India. *Genome Announcements*, 4(1), 1704–1715.
- Aghaie, E., Pazouki, M., Hosseini, M. R., Ranjbar, M., & Ghavipanjeh, F. (2009). response surface methodology (RSM) analysis of organic acid production for kaolin beneficiation by *Aspergillus niger*. *Chemical Engineering Journal*, 147, 245–251.
- Aguilera, F., Mendez, J., Pasaro, E., & Laffon, B. (2010). Review on the effects of exposure to spilled oils on human health. *Journal of Applied Toxicology*, 30(4), 291–301.
- Aislabie, J., McLeod, M., & Fraser, R. (1998). Potential of biodegradation of hydrocarbons in soil from the Ross Dependency, Antarctica. *Applied Microbiology and Biotechnology*, 49, 210–214.
- Aislabie, J., Foght, J., & Saul, D. (2000). Aromatic hydrocarbon-degrading bacteria from soil near Scott Base, Antarctica. *Polar Biology*, 23, 183–188.
- Aislabie, J., Balks, M., Foght, J., & Waterhouse, E. (2004). Hydrocarbon spills on Antarctic soils: effects and management. *Environmental Science and Technology*, 38(5), 1266–1274.
- Aislabie, J., Saul, D., & Foght, J. (2006). Bioremediation of hydrocarbon-contaminated polar soils. *Extremophiles*, 10, 171–179.
- Akpe, A. R., Ekundayo, A. O., & Esumeh, F. I. (2013). Degradation of crude oil by bacteria: a role for plasmid borne genes. *Global Journal of Science Frontier Research Biological Science*, 13(6), 1-7.
- Alexander, M. (1999). *Biodegradation and Bioremediation*. 2nd ed. New York: Academic Press.

- Alkan, C., Sajadian, S., & Eichler, E. E. (2011). Limitations of next-generation genome sequence assembly. *Nature Methods*, 8(1), 61-65.
- Altschul, S. F., Gish, W., Miller, W., Myers, E. W., & Lioman, D. J. (1990). Basic local alignment search tool. *Journal of Molecular Biology*, 215, 403-410.
- Alvarez, L. M. M., Lo Balbo, A., Mac Cormack, W. P., & Ruberto, L. A. M. (2015). Bioremediation of a petroleum hydrocarbon-contaminated Antarctic soil: optimisation of a biostimulation strategy using response surface methodology (RSM). *Cold Regions Science and Technology*, 119, 61-67.
- Anderson, M. J., & Whitcomb, P. J. (2004). RSM Simplified: Optimizing Processes using response surface methods for design of experiments. *Business and Economics*, CRC Press, 304 pp.
- Ansorge, W., Sproat, B. S., Stegemann, J., & Schwager, C. (1986). A non-radioactive automated method for DNA sequence determination. *Journal of Biochemical and Biophysical Methods*, 13(6), 315-323.
- Ansorge, W., Sproat, B., Stegemann, J., Schwager, C., & Zenke, M. (1987). Automated DNA sequencing: ultrasensitive detection of fluorescent bands during electrophoresis. *Nucleic Acids Research*, 15(11), 4593-4602.
- Armstrong, B. G., Hutchinson, E., Unwin, J., & Fletcher, T. (2004). Lung cancer risk after exposure to polycyclic aromatic hydrocarbons; a review and meta-analysis. *Environmental Health Perspectives*, 112(9), 970-978.
- Atlas, R. M. (1991). Microbial hydrocarbon degradation - Bioremediation of oil spills. *Journal of Chemical Technology and Biotechnology*, 52, 149-156.
- Atlas, R. M. (1986). Fate of petroleum pollutants in Arctic ecosystems. *Water Science and Technology*, 18(2), 59-67.
- Atlas, R. M., & Bartha, R. (1998). *Microbial ecology: fundamentals and applications*. Benjamin/Cummings Publishing Company, pp 281-324.
- Atlas, R., & Bragg, J. (2009). Bioremediation of marine oil spills: when and when not - the Exxon Valdez experience. *Microbial Biotechnology*, 2(2), 213-221.
- ATCM (1998). *Final report of the twenty-second Antarctic Treaty Consultative Meeting*, Antarctic Treaty Consultative Meeting: Tromso, Norway.
- ATSDR (Agency for Toxic Substances and Disease Registry) (1995). Toxicological profile for polycyclic aromatic hydrocarbons (PAHs). Atlanta, GA, US Department of Health and Human Services, Public Health Service.
- ATSDR (Agency for Toxic Substances and Disease Registry) (1999). Toxicological profile for total petroleum hydrocarbons (TPH). Atlanta, GA, US Department of Health and Human Services, Public Health Service.

- Austin, R. N., Chang, H-K., Zylstra, G. J., & Groves, J. T. (2000). The non-heme diiron alkane monooxygenase of *Pseudomonas oleovorans* (AlkB) hydroxylates via a substrate radical intermediate. *Journal of American Chemical Society*, 122, 11747–11748.
- Aziz, R. K., Bartels, D., Best, A. A., DeJongh, M., Disz, T., Edwards, R. A., Formsma, K., et al. (2008). The RAST server: rapid annotations using subsystems technology. *BMC Genomics*, 9,75. <http://dx.doi.org/10.1186/1471-2164-9-75>.
- Bader, R. F. W., Tang, T. H., Tal, Y., & Biegler-Koenig, F. W. (1982). Molecular structure and its change: hydrocarbons. *Journal of the American Chemical Society*, 104(4), 940-945. doi: 10.1021/ja00368a003.
- Balasubramanian, S. (2011). Sequencing nucleic acids: from chemistry to medicine. *Chem Commun (Camb)*, 47(26), 7281–7286.
- Bankevich, A., Nurk, S., Antipov, D., Gurevich, A. A., Dvorkin, M., Kulikov, A. S., Lesin, V. M., et al. (2012). SPAdes: a new genome assembly algorithm and its application to single cell sequencing. *Journal of Computational Biology*, 19(5), 455-477. doi: 10.1089/cmb2012.0021..
- Baraniecki, C. A., Aislabilie, J., & Foght, J. M. (2002). Characterization of *Sphingomonas* sp. Ant 17, an aromatic hydrocarbon-degrading bacterium isolated from Antarctic soil. *Microbial Ecology*, 43, 22–54.
- Baş, D., & Boyaci, I. H. (2007). Modelling and optimization I: usability of response surface methodology. *Journal of Food Engineering*, 78(3), 836-845. doi: 10.1016/j.jfoodeng.2005.11.024.
- Bej, A. K., Saul, D., & Aislabilie, J. (2000). Cold-tolerant alkane-degrading *Rhodococcus* species from Antarctica. *Polar Biology*, 23, 100–105.
- Bell, K. S., Philip, J. C., Aw, D. W., & Christofi, N. (1998). The genus *Rhodococcus*. *Journal of Applied Microbiology*, 85, 195-210. <https://doi.org/10.1046/j.1365-2672.1998.00525.x>.
- Bosset, I. & Bartha, R. (1984). The fate of petroleum in soil ecosystems. In Atlas, R. M. (Ed), *Petroleum Microbiology* (pp. 435-476). New York, NY: Macmillan Publishing Company.
- Belhaj, A., Desnoues, N., & Elmerich, C. (2002). Alkane biodegradation in *Pseudomonas aeruginosa* strains isolated from a polluted zone: identification of *alkB* and *alkB*-related genes. *Research in Microbiology*, 153(6), 339–344.
- Bentley, D. R. (2006). Whole genome re-sequencing. *Current Opinion in Genetics and Development*, 16(6), 545-552.

- Bentley, D. R., Balasubramanian, S., Swerdlow, H. P., Smith, G. P., Milton, J., Brown, C. G., Hall, K. P., et al. (2008). Accurate whole human genome sequencing using reversible terminator chemistry. *Nature*, 456(7218), 53-59. doi: 10.1038/nature07517.
- Bockheim, J. G., & McLeod, M. (2006). Soil formation in Wright Valley, Antarctica since the late Neogene. *Geoderma*, 137(1), 109–116. <https://doi.org/10.1016/j.geoderma.2006.08.028>
- Box, G. E. P., & Wilson, K. B. (1951). On the Experimental Attainment of Optimum Conditions. *Journal of the Royal Statistical Society. Series B (Methodological)*, 13(1), 1–45.
- Brennerova, M. V., Josefiova, J., Brenner, V., Pieper, D. H., & Junca, H. (2009). Metagenomics reveals diversity and abundance of *meta*-cleavage pathways in microbial communities from soil highly contaminated with jet fuel under air-sparging bioremediation. *Environmental Microbiology*, 11(9), 2216–2227.
- Buchanan, B. B. (1991). Regulation of CO₂ assimilation in oxygenic photosynthesis: the ferrodoxin thioredoxin system: perspective on its discovery, present status, and future development. *Archive of Biochemistry and Biophysics*, 288, 1-9.
- Bushnell, L. D., & Haas, H. F. (1941). The utilization of certain hydrocarbons by microorganisms. *Journal of Bacteriology*, 41(5), 653-673.
- Caporaso, J. G., Lauber, C. L., Walters, W. A., Berg-Lyons, D., Huntley, J., Fierer, N., Owens, S. M., et al. (2012). Ultra-high-throughput microbial community analysis on the Illumina HiSeq and MiSeq platforms. *The ISME Journal*, 6(8), 1621-1624.
- Cappuccino, J. G., & Sherman, N. (2005). Microbiology: a laboratory manual (7th ed.) San Francisco: Pearson/Benjamin Cummings
- Camacho, C., Coulouris, G., Avagyan, V., Ma, N., Papadopoulos, J., Bealer, K., & Madeen, T. L. (2009). BLAST+: architecture and applications. *BMC Bioinformatics*, 10(421). doi: 10.1186/1471-2105-10-421.
- Camenzuli, D., Freidman, B. L., Statham, T. M., Mumford, K. A., & Gore, D. B. (2013). On-site and *in situ* remediation technologies applicable to petroleum hydrocarbon contaminated sites in the Antarctic and Arctic. *Polar Research*, 33, doi: <http://dx.doi.org/10.3402/polar.v33.21522>.
- Camenzuli, D. & Freidman, B. L. (2015). On-site and *in situ* remediation technologies applicable to petroleum hydrocarbon contaminated sites in the Antarctic and Arctic. *Polar Research*, 34, <http://dx.doi.org/10.3402/polar.v34.24492>.
- Campbell, I. B., Claridge, G. G. C., Campbell, D. I., & Balks, M. R. (1998). The soil environments of the McMurdo Dry Valleys, Antarctica, in ecosystem dynamics in a polar desert: the McMurdo Dry Valleys, Antarctica. In Priscu, J. C., *American Geophysical Union*, Washington , D. C. doi: 10.1029/AR072p0297.

- Canard, B., & Sarfati, R. S. (1994). DNA polymerase fluorescent substrates with reversible 3'-tags. *Gene*, 148(1), 1-6.
- Carmichael, L. M., & Pfaender, F. K. (1997). The effect of inorganic and organic supplements on the microbial degradation of phenanthrene and pyrene in soils. *Biodegradation*, 8(1), 1-13.
- Case, R. J., Boucher, Y., Dahllof, I., Homstrom, C., Doolittle, W. F., & Kjelleberg, S. (2007). Use of 16S rRNA and *ropB* genes as molecular markers for microbial ecology studies. *Applied and Environmental Microbiology*, 73(1), 278-288.
- Chaineau, C. H., Morel, J., Dupont, J., Bury, E., & Oudot, J. (1999). Comparison of the fuel oil biodegradation potential of hydrocarbon-assimilating microorganisms isolated from a temperate agricultural soil. *Science of Total Environment*, 227(2-3), 237–247.
- Chaineau, C. H., Rougeux, G., Yeremian, C., & Oudot, J. (2005). Effect of nutrient concentration on the biodegradation of crude oil and associated microbial populations in the soil. *Soil Biology and Biochemistry*, 20, 1-8.
- Chuvilin, E. M., Naletova, N. S., Miklyeva, E. C., Kozlova, E. V., & Instanes, A. (2001). Factors affecting spreadability and transportation of oil in regions of frozen ground. *Polar Record*, 37, 229–238.
- Compeau, G., Al-achi, B. J., Platsouka, E., & Levy, S. B. (1988). Survival of rifampin-resistant mutants of *Pseudomonas fluorescens* and *Pseudomonas putida* in soil systems. *Applied and Environmental Microbiology*, 54(10), 2432-2438.
- Cookson, J. T. (1995). Bioremediation engineering design and application. McGraw Hill, New York.
- Cooley, R. B., Dubbels, B. L., Sayavedra-Soto, L. A., Bottomley, J. & Arp, D. J. (2009). Kinetic characterization of the soluble butane monooxygenase from *Thauera butanivorans*, formerly '*Pseudomonas butanovora*'. *Microbiology*, 155(6), 2086-2096.
- Coulon, F., & Delille, D. (2003). Effects of biostimulation on growth of indigenous bacteria in sub-Antarctic soil contaminated with oil hydrocarbons. *Oil and Gas Science and Technology*, 58(4), 469-479.
- Coulon, F., Pelletier, E., St. Louis, R., Gourhant, L. & Delille, D. (2004). Degradation of petroleum hydrocarbons in two sub-Antarctic soils: influence of an oleophilic fertilizer. *Environmental Toxicology and Chemistry*, 23(8), 71893-1901.
- Cripps, G. C., & Priddle, J. (1991). Hydrocarbons in the Antarctic marine environment. *Antarctic Science*, 3(3), 233-250. doi: 10.1017/S0954102091000299
- Curtosi, A., Pelletier, E., Vodopivec, C. L., & Mac Cormack, W. P. (2007). Polycyclic aromatic hydrocarbons in soil and surface marine sediments near Jubany Station (Antarctica) Role of permafrost as a low-permeability barrier. *Science of the Total Environment*, 383, 193–204.

- Czitrom, V. (1999). One-Factor-at-a-Time versus Designed Experiments. *The American Statistician*, 53(2), 126–131. <https://doi.org/10.1080/00031305.1999.10474445>
- Das, N. & Chandran, P. (2011). Microbial degradation of petroleum hydrocarbon contaminants: an overview. *Biotechnology Research International*, doi: 10.4061/2011/941810.
- Date, A. W. (2011). *Analytical Combustion: With Thermodynamics, Chemical Kinetics and Mass Transfer* (Google eBook). Cambridge University Press. ISBN 1-107-00286-9.
- Dawson, R. M. C., Elliott, D. C., Elliott, W. H., & Jones, K. M. (1986). Data for biochemical research. *Oxford Science*, Oxford.
- Dear, P. H. (2005). Genome Mapping. eLS. doi: 10.1038/npg.els.0005353.
- de Carvalho, C. C.R., & da Fonseca, M. M. R. (2005). Degradation of hydrocarbons and alcohols at different temperatures and salinities by *Rhodococcus erythropolis* DCL14. *FEMS Microbiology Ecology*, 51(3), 389-399.
- de Carvalho, C. C. R., da Cruz, A. A. R. L., Pons, M-N., Pinheiro, H. M. R. V., Cabral, J. M. S., da Fonseca, M. M. R., Ferreira, B. S. & Fernandes, P. (2004). *Mycobacterium* sp., *Rhodococcus erythropolis*, and *Pseudomonas putida* behaviour in the presence of organic solvents. *Microscopy Research and Technique*, 64(3), 215-222.
- Decker, A., & Solomon, E. I. (2005). Dioxygen activation by copper, heme and non-heme iron enzymes: comparison of electronic structure and reactivities. *Current Opinion in Chemical Biology*, 9(2), 152-163.
- De Domenico, M., Lo Giudice, A., Michaud, L., Saitta, M., & Bruni, V. (2004). Diesel and PCB-degrading psychrotrophic bacteria isolated from Antarctic seawaters (Terra Nova Bay, Ross Sea). *Polar Research*, 23(2), 141–146.
- Delille, D., & Coulon, F. (2008). Comparative mecosm study of biostimulation efficiency in two different oil-amended sub-Antarctic soils. *Microbial Ecology*, 56(2), 243-252.
- Delille, D., Pelletier, E., Delille, B. & Coulon, F. (2003). Effect of nutrient enrichments on the bacterial assemblage of Antarctic soils contaminated by diesel or crude oil. *Polar Research*, 39, 1-10.
- Dennett, G. V., & Blamey, J. M. (2016). A new thermophilic nitrilase from an Antarctic hyperthermophilic microorganism. *Frontiers in Bioengineering and Biotechnology*, 4(5). doi: 10.3389/fbioe.2016.00005.
- Desjardins, P., & Conklin, C. (2010). NanoDrop microvolume quantitation of nucleic acids. *Journal of Visualized Experiments*, 45, e2565, 1-4.

- Dias, R. L., Ruberto, L., Hernandez, E., Vazquez, S. C., Lo Balbo, A., Del Panno, M. T., et al. (2012). Bioremediation of an aged diesel oil-contaminated Antarctic soil: evaluation of the "on site" biostimulation strategy using different nutrient sources. *International Biodeterioration and Biodegradation*, 75, 96–103.
- Diks, R. M. M., Ottengraf, S. P. P., & van den Oever, A. H. C. (1994). The influence of NaCl on the degradation rate of dichloroethane by *Hyphomicrobium* sp. *Biodegradation*, 5, 129-141.
- Dojka, M. A., Hugenholtz, P., Haack, S. K., & Norman, R. P. (1998). Microbial diversity in a hydrocarbon- and chlorinated-solvent-contaminated aquifer undergoing intrinsic bioremediation. *Applied and Environmental Microbiology*, 64(10), 3869–3877.
- Donkor, E. S., Dayie, N. T. K. D., & Adiku, T. K. (2014). Bioinformatics with basic local alignment search tool (BLAST) and fast alignment (FASTA). *Journal of Bioinformatics abd Sequence Analysis*, 6(1), 1–6.
- Durán, N., & Esposito, E. (2000). Potential applications of oxidative enzymes and phenoloxidase-like compounds in wastewater and soil treatment: a review. *Applied Catalysis B: Environmental*, 28, 83-99.
- Eddy, S. R. (1998). Profile Hidden Markov Models. *Bionformatics*, 14, 755-763.
- Emtiazi, G., Mridamadian, S., & Habibi, M. H. (2005). Instability of petroleum oil degradation by induction of mutation in 45kb and 60kb plasmids. *International Journal of Environmental Studies*, 62(4), 467–472.
- Engelbrektson, A., Kunin, V., Wrighton, K. C., Zvenigorodsky, N., Chen, F., Ochman, H., & Hugenholtz, P. (2010). Experimental factors affecting PCR-based estimates of microbial species richness and evenness. *The ISME Journal*, 4(5), 642–647. <https://doi.org/10.1038/ismej.2009.153>
- Eweis, J. B., Ergas, S. J., Chang, D. P. Y., & Schroeder, E. D. (1998). Bioremediation Principles, *McGraw-Hill*, Boston.
- Fasan, R., Mehareenna, Y. T., Snow, C. D., Poulos, T. L., & Arnold, F. H. (2008). Evolutionary history of a specialized P450 propane monooxygenase. *Journal of Molecular Biology*, 383(5), 1069-1080.
- Fedorco, M., Romieu, A., Williams, S., Lawrence, I., & Turcatti, G. (2006). BTA, a novel reagent for DNA attachment on glass and efficient generation of solid-phase amplified DNA colonies. *Nucleic Acids Research*, 34(3).
- Felsenstein, J. (1985). Confidence limits on the phylogenies: an approach using the bootstrap. *Evolution*, 39(4), 783-791.

- Feng, L., Wang, W., Cheng, J., Ren, Y., Zhao, G., Gao, C., et al. (2007). Genome and proteome of long-chain alkane degrading *Geobacillus thermodenitrificans* NG80-2 isolated from a deep-subsurface oil reservoir. *Proceedings of the National Academy of Sciences of USA*, 104(13), 5602-5607.
- Ferguson, S. H., Franzmann, P. D., Revill, A. T., Snape, I., & Rayner, J. L. (2003). The effects of nitrogen and water on mineralisation of hydrocarbons in diesel-contaminated terrestrial Antarctic soils. *Cold Regions Science and Technology*, 37, 197-212.
- Filler, D. M., Reynolds, C. M., Snape, I., Dagulis, A. J., Barnes, D. L., & Williams, P. J. (2006). Advances in engineered remediation for use in the Arctic and Antarctic. *Polar Record*, 42, 111-120.
- Fleischmann, R., Adams, M., White, O., Clayton, R., Kirkness, E., Kerlavage, A., Bult, C., et al. (1995). Whole-genome random sequencing and assembly of *Haemophilus influenzae* Rd. *Science*, 269(5223), 496-512. doi: 10.1126/science.7542800.
- Fletcher, G. L., Hew, C. L., & Davies, P. L. (2001). Antifreeze proteins of teleost fishes. *Annual Review of Physiology*, 63, 359-390.
- Florou, A. B., Prodromidis, M. I., Tzouwara-Karayanni, S. M., & Karayannidis, M. I. (2000). Fabrication and voltammetric study of lanthanum 2,6-dichlorophenolindophenol chemically modified screen printed electrodes: application for the determination of ascorbic acid. *Analytica Chimica Acta*, 423, 107-114.
- Fox, G. E., Magrum, L. J., Balch, W. E., Wolfe, R. S., & Woese, C. R. (1977). Classification of methanogenic bacteria by 16S ribosomal RNA characterization. *Proceedings of the National Academy of Science USA*, 74(10), 4537-4541.
- Franzmann, P. D. (1996). examination of Antarctic prokaryotic diversity through molecular comparisons. *Biodiversity and Conservation*, 85(11), 1295–1305.
- Fritsche, W., & Hofrichter, M. (2008). Aerobic degradation by microorganisms. *Biotechnology Set, Second Edition*, pp. 144–167.
- Gallagher, P. E., Weiss, R. B., Brent, T. P., & Duker, N. J. (1989). Wavelength dependence of DNA incision by a human ultraviolet endonuclease. *Photochemistry and Photobiology*, 49(3), 363-367. doi: 10.1111/j.1751-1097.1989.tb04120.x.
- Gallego, J. L. R., Loredo, J., Llamas, J. F., Vazquez, F., & Sanchez, J. (2001). Bioremediation of diesel-contaminated soils: evaluation of potential *in situ* techniques by study of bacterial degradation. *Biodegradation*, 12, 325-335.

- Gesheva, V., Stackebrandt, E. & Vasileva-Tonkova, E. (2010). Biosurfactant production by halotolerant *Rhodococcus fascians* from Casey Station, Wilkes Land, Antarctica. *Current Microbiology*, 61, 112–117.
- Gibson, G., & Muse, S. V. (2009). A Primer of Genome Science (3rd ed). Sinauer Associates, pp 84.
- Gibson, D. T., & Parales, R. E. (2000). Aromatic hydrocarbon dioxygenases in environmental biotechnology. *Current Opinion in Biotechnology*, 11(3), 236–243.
- Gibson, J., Shokralla, S., Porter, T. M., King, I., van Konynenburg, S., Janzen, D. H., Hallwachs, W., & Hajibabaei, M. (2014). Simultaneous assessment of the macrobiome and microbiome in a bulk sample of tropical arthropods through DNA metasystemics. *Proceedings of the National Academy of Sciences of USA*, 111(22), 8007-8012.
- Giorgio, M., Trinei, M., Migliaccio, E., & Pelicci, P. G. (2007). Hydrogen peroxide: a metabolic by-product or a common mediator of ageing signals? *Nature Reviews Molecular Cell Biology*, 8, 722-728. doi: 10.1038/nrm2240.
- Goodsell, D. S. (2009). Molecule of the month: antifreeze proteins. *Worldwide Protein Data Bank*, New Jersey, USA.
- Goordial, J., Raymond-Bouchard, I., Ronholm, J., Shapiro, N., Woyke, T., Whyte, L., et al. (2015). Improved-high-quality draft gene sequence of *Rhodococcus* sp. JG-3, a eurypsychrophilic Actinobacteria from Antarctic Dry Valley permafrost. *Standard in Genomic Sciences*, 10(61), doi: 10.1186/s40793-015-0043-8.
- Greer, C. W., Whyte, L. G., Niederberger, T. D., & Timmis, K. N. (2010). Microbial communities in hydrocarbon contaminated temperate, tropical, alpine, and polar soils. *Handbook of Hydrocarbon and Lipid Microbiology*. Berlin Springer, 2313-2328.
- Greenshields, J. B., & Rossini, F. D. (1958). Molecular structure and properties of hydrocarbons and related compounds, *The Journal of Physical Chemistry*, 62(3), 271-280.
- Gao, B., & Gupta, R. S. (2012). Phylogenetic framework and molecular signatures for the main clades of the phylum Actinobacteria. *Microbiology and Molecular Biology Reviews*, 76(1), 66–112.
- Goody, R. S. (2013). How Bacteria Choose Phosphate. *Angewandte Chemie International Edition*, 52(9), 2406–2407. <https://doi.org/10.1002/anie.201209376>
- Haas, D., & Defago, G. (2005). Biological control of soil-borne pathogens by fluorescent pseudomonads. *Nature Reviews: Microbiology*, 3(4), 307-319.

- Haas, D., & Keel, C. (2003). Regulation of antibiotic production in root-colonizing *Pseudomonas* spp. and relevance for biological control of plant disease. *Annual Review of Phytopathology*, 41, 117-153.
- Haddock, J. D. (2010). Aerobic degradation of aromatic hydrocarbon: enzyme structures and catalytic mechanisms. In Timmis, K. N. (ed.), *Handbook of Hydrocarbon and Lipid Microbiology*, doi: 10.1007/978-3-540-77587-4_74, Springer-Verlag Berlin Heidelberg.
- Hamamura, N., Storfa, R. T., Semprini, L. & Arp, D. J. (1999). Diversity in butane monooxygenase among butane-grown bacteria. *Applied and Environmental Microbiology*, 65(10), 4586-4593.
- Hanlon, D. W., & Ordal, G. W. (1994). Cloning and characterization of genes encoding methyl-accepting chemotaxis proteins in *Bacillus subtilis*. *Journal of Biological Chemistry*, 269(19), 14038-14046.
- Hanlon, D. W., Carpenter, P. B., & Ordal, G. W. (1992). Influence of attractants and repellents on methyl group turnover on methyl-accepting chemotaxis proteins of *Bacillus subtilis* and role of CheW. *Journal of Bacteriology*, 174(13), 4218-4222.
- Hanson, K. G., Desai, J. D., & Desai, A. J. (1993). A rapid and simple screening technique for a potential crude oil degrading microorganism. *Journal of Chemical Technology and Biotechnology*, 7, 745-748.
- Harayama, S., Kok, M. & Neidle, E. L. (1992). Functional and evolutionary relationships among diverse oxygenases. *Annual Review of Microbiology*, 46, 565–601.
- Harayama, S., Kishira, H., Kasai, Y. & Shutsubo, K. (1999). Petroleum biodegradation in marine environments. *Journal of Molecular Microbiology and Biotechnology*, 1(1), 63-70.
- Harwood, C. S., & Ornston, L. N. (1984). TOL plasmid can prevent induction of chemotactic responses to aromatic acids. *Journal of Bacteriology*, 160(2), 797-800.
- Hathaway, L. J., Brugger, S., Martynova, A., Aebi, S., & Mühlmann, K. (2007). Use of the Agilent 2100 Bioanalyzer for Rapid and Reproducible Molecular Typing of *Streptococcus pneumoniae*. *Journal of Clinical Microbiology*, 45(3), 803–809. <https://doi.org/10.1128/JCM.02169-06>
- Hazan, R., Que, Y.-A., Maura, D., & Rahme, L. G. (2012). A method for high throughput determination of viable bacteria cell counts in 96-well plates. *BMC Microbiology*, 12, 259.
- Hazen, T. C. (1994). Chemotactic selection of pollutants degrading soil bacteria, In U.S. Patent 5324661.

- Hazen, T. C., & Lopez-de-Victoria, G. (1994). Method of degrading pollutants in soil, In U.S. Patent 5236703
- Head, I. M., & Swannell, R. P. (1999). Bioremediation of petroleum hydrocarbon contaminants in marine habitats. *Current Opinion in Biotechnology*, 10(3), 234-239.
- Heather, J. M., & Chain, B. (2016). The sequence of sequencers: the history of sequencing DNA. *Genomics*, 107(1), 1-8.
- Heider, J., Spormann, A. M., Beller, H. R., & Widdel, F. (1998). Anaerobic bacterial metabolism of hydrocarbons. *FEMS Microbiology Reviews*, 22(5), 459–473.
- Hunkapiller, T., Kaiser, R. J., Koop, B. F., & Hood, L. (1991). Large-scale and automated DNA sequence determination. *Science*, 254(5028), 59-67.
- Hyatt, D., Chen, G. L., Locascio, P. F., Land, M. L., Larimer, F. W., & Hauser, L. J. (2010). Prodigal: prokaryotic gene recognition and translation initiation site identification. *BMC Bioinformatics*, 11(119). doi: 10.1186/1471-2105-11-119.
- Hyman, E. D. (1988). A new method of sequencing DNA. *Analytical Biochemistry*, 174(2), 423–436.
- ITRC (Interstate Technology and Regulatory Council). (2014). *Petroleum Vapor Intrusion; Fundamentals of Screening, Investigation, and Management*. PVI-1. Washington, D.C.: Interstate Technology and Regulatory Council, Petroleum Vapor Intrusion Team. www.itrcweb.org/PetroleumVI-Guidance.
- Ismail, W., & Gescher, J. (2012). Epoxy coenzyme a thioester pathway for degradation of aromatic compounds. *Applied and Environmental Microbiology*, 78(15), 5043–5051.
- Jain, P. K., Gupta, V. K., Gaur, R. K., Lowry, M., Jaroli, D. P., & Chauhan, U. K. (2011). Bioremediation of petroleum oil contaminated soil and water. *Research Journal of Environmental Toxicology*, 5, 1–26. doi: 10.3923/rjet.2011.1.26.
- Janda, J. M., & Abbott, S. L. (2007). 16S rRNA gene sequencing for bacterial identification in diagnostic laboratory: pluses, perils, and pitfalls. *Journal of Clinical Microbiology*, 45, 2761–2764.
- Jansson, M. (1988). Phosphate uptake and utilization by bacteria and algae. *Hydrobiologia*, 170(1), 177–189. <https://doi.org/10.1007/BF00024904>
- Jesus, H. E., Peixoto, R. S., & Rosado, A. S. (2015). Bioremediation in Antarctic soils. *Environmental Biotechnology*, 6(6), 1-11.
- John, R. C., & Okpokwasili, G. C. (2012). Crude oil-degradation and plasmid profile of nitrifying bacteria isolated from oil-impacted mangrove sediment in the Niger Delta of Nigeria. *Bulletin of Environmental Contamination and Toxicology*, 88, 1020-1026.

- Johnson, E. L., & Hyman, M. R. (2006). Propane and *n*-butane oxidation by *Pseudomonas putida* GPo1. *Applied and Environmental Microbiology*, 72(1), 950-952.
- Johnson, G. R., & Olsen, R. H. (1997). Multiple pathways for toluene degradation in *Burkholderia* sp. strain JS150. *Applied and Environmental Microbiology*, 63, 4047-4052.
- Jones, O. A. H., Spurgeon, D. J., Svendsen, C., & Griffin, J. L. (2008). A metabolomics based approach to assessing the toxicity of the polycyclic aromatic hydrocarbon pyrene to the earthworm *Lumbricus rubellus*. *Chemosphere*, 71(3), 601-609.
- Joseph, S. J., Hugenholtz, P., Sangwan, P., Osborne, C. A., & Janssen, P. H. (2003). Laboratory cultivation of widespread and previously uncultured soil bacteria. *Applied Environmental Microbiology*, 69(12), 7210-7215.
- Jouanneau, Y., Meyer, C., Jakoncic, J., Stojanoff, V., & Gaillard, J. (2006). Characterization of a naphthalene dioxygenase endowed with an exceptionally broad substrate specificity toward polycyclic aromatic hydrocarbons. *Biochemistry*, 45, 12380-12391.
- Jukes, T. H., & Cantor, C. R. (1969). *Evolution of protein molecules*. In: Munro, H. N. (Ed). *Mammalian Protein Metabolism III*. Academic Press, New York, pp 21-132.
- Kaczorek, E., & Olszanowski, A. (2011). Uptake of hydrocarbon by *Pseudomonas fluorescens* (P1) and *Pseudomonas putida* (K1) strains in the presence of surfactants: a cell surface modification. *Water, Air and Soil Pollution*, 214(1-4), 451-459.
- Kambara, H., Nishikawa, T., Katayama, Y., & Yamaguchi, T. (1988). Optimization of parameters in a DNA sequenator using fluorescence detection. *Nature Biotechnology*, 6, 816-821.
- Kalman, B. & Johnson, R. (2007). *Endangered Penguins*: New York: Crabtree Publishing and Cooperation.
- Kauppi, S., Sinkkonen, A., & Romantschuk, M. (2011). Enhancing bioremediation of diesel-fuel-contaminated soil in a boreal climate: comparison of biostimulation and bioaugmentation. *International Biodeterioration and Biodegradation*, 65(2), 359-368. doi: 10.1016/j.ibiod.2010.10.011.
- Kennicutt, M. C. (1990). Oil spillage in Antarctica: initial report of the National Science Foundation-sponsored Quick Response Team on the grounding of the Bahia Paraiso. *Environmental Science and Technology*, 24(5), 620-624.
- Kennicutt, M. C., McDonald, T. J., Denoux, G. J., & McDonald, S. J. (1992). Hydrocarbon contamination on the Antarctic Peninsula: I. Arthur harbor-subtidal sediments. *Marine Pollution Bulletin*, 24(10), 499-506. [https://doi.org/10.1016/0025-326X\(92\)90474-K](https://doi.org/10.1016/0025-326X(92)90474-K).

- Kim, K., Jahan, S. A., Kabir, E., & Brown, R. J. C. (2013). A review of airborne polycyclic aromatic hydrocarbons (PAHs) and their human health effects. *Environmental International*, 60, 71-80.
- Kinawy, A. A. (2009). Impact of gasoline inhalation on some neurobehavioural characteristics of male rats. *BMC Physiology*, 9(21). doi: 10.1186/1472-6793/9/21
- Knothe, G. (2010). Biodiesel and renewable diesel: a comparison. *Progress in Energy and Combustion Sciences*, 36, 364-373.
- Komukai-Nakamura, S., Sugiura, K., Yamauchi-Inomata, Y., Toki, H., Venkateswaran, K., Yamamoto, S., Tanaka, H., & Harayama, S. (1996). Construction of bacterial consortia that degrade Arabian light crude oil. *Journal of Fermentation and Bioengineering*, 382(6), 570-574.
- Konidari, C. N., Tzouware, S. M., Bowman, L. E., & Karayannis, M. I. (1992). Kinetic and mechanistic study of the reaction of 2,6-dichlorophenolindophenol and cysteine. *Talanta*, 39, 863-868.
- Kosina, M., Barták, M., Mašlaňová, I., Pascutti, A. V., Sedo, O., Lexa, M., & Sedláček, I. (2013). *Pseudomonas proskeii* sp. nov., a novel psychrotrophic bacterium from Antarctica. *Current Microbiology*, 67(6), 637-646. <https://doi.org/10.1007/s00284-013-0406-6>
- Kotani, T., Yamamoto, T., Yurimoto, H., Sakai, Y., & Kato, N. (2003). Propane monooxygenase and NAD⁺ dependent secondary alcohol dehydrogenase in propane metabolism by *Gordonia* sp. strain TY-5. *Journal of Bacteriology*, 185, 7120-7128.
- Krastanov, A., Alexieva, Z., & Yemendzhiev, H. (2013). Microbial degradation of phenol and phenolic derivatives. *Engineering in Life Sciences*, 13(1), 76-87.
- Kuhn, E., Bellicanta, G. S., & Pellizari, V. H. (2009). New alk genes detected in Antarctic marine sediments. *Environmental Microbiology*, 11(3), 669–673.
- Kuo, C. Y., Hsu, Y. W. & Lee, H. S. (2003). Study of human exposure to particulate PAHs using personal air samplers. *Archive of Environmental Contamination and Toxicology*, 44, 454–459.
- Labinger, J. A., & Bercaw, J. E. (2002). Understanding and exploiting C-H bond activation. *Nature*, 417(6888), 507–514.
- Laczi, K., Kis, A., Horvath, B., Maroti, G., Hegedus, B., Perei, K., & Rakheley, G. (2015). Metabolic responses of *Rhodococcus erythropolis* PR4 grown on diesel oil and various hydrocarbons. *Applied Microbiology and Biotechnology*, 99(22), 9745-9759.

- Lagesen, K., Hallin, P., Rodland, E. A., Staerfeldt, H. H., Rognes, T., & Ussery, D. W. (2007). RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Research*, 35(9), 3100-3108.
- Laslett, D. & Bjorn, C. (2004). ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. *Nucleic Acids Research*, 32(1), 11-16.
- Laurence, M., Hatzis, C., & Brash, D. E. (2014). Common contaminants in next-generation sequencing that hinder discovery of low-abundance microbes. *PLoS One*. 9(5), e97876. doi: 10.1371/journal.pone.0097876.
- Leahy, J. G., Batchelor, P. J., & Morcomb, S. M. (2003). Evolution of the soluble diiron monooxygenases. *FEMS Microbiology Reviews*, 27(4), 449–714.
- Lee, C-H., Liang, S., Haumann, T., Boese, R., & de Meijere, A. (1993). *p*-[3^{2,5⁶}] octahedrane, the (CH)₁₂ hydrocarbon with *D*_{3d} symmetry. *Angewandte Chemie International Edition*, 32(4), 559-561.
- Liang, J-L., JiangYang, J-H., Nie, Y. & Wu, X-L. (2016). Regulation of alkane hydroxylase CYP153 gene in a gram-positive alkane-degrading bacterium, *Dietzia* sp. strain DQ12-45-1b. *Applied and Environmental Microbiology*, 82(2), 608–619.
- Likhoshvay, A., Lomakina, A., & Grachev, M. (2014). The complete *alk* sequences of *Rhodococcus erythropolis* from Lake Baikal. *SpringerPlus*, 39(621). doi: 10.1186/2193-1801-3-621.
- Lo Giudice, A., Bruni, V., De Domenico, M. & Michaud, L. (2010). Psychrophiles - cold-adapted hydrocarbon-degrading microorganisms. In Timmis, K. N. (Ed), *Handbook of Hydrocarbon and Lipid Microbiology*, doi: 10.1007/978-3-540-77587-4, 1897-1921.
- Loman, N. J., Constantinidou, C., Chan, J. Z. M., Halachev, M., Sergeant, M., Penn, C. W., Robinson, E. R., & Pallen, M. J. (2012). High-throughput bacterial genome sequencing: an embarrassment of choice, a world of opportunity. *Nature Reviews Microbiology*, 10, 499-606.
- Long, J., Ma, J., Luo, C., Mo, X., Sun, L., Zang, W., & Liu, J. (2009). Comparison of two methods for assaying complex I activity in mitochondrial isolated from rat liver, brain and heart. *Life Science*, 85, 276-280.
- Luckey, J. A., Drossman, H., Kosticjka, A. J., Mead, D. A., D'Cunha, J., Norris, T. B., & Smith, L. M. (1990). High speed DNA sequencing by capillary electrophoresis. *Nucleic Acids Research*, 18(15), 4417–4421.
- Ludwig, B., Akundi, A., & Kendall, K. (1995). A long-chain secondary alcohol dehydrogenase from *Rhodococcus erythropolis* ATCC 4277. *Applied and Environmental Microbiology*, 61(10), 3729-3733.

- Lugtenberg, B., & Kamilova, F. (2009). Plant-growth-promoting rhizobacteria. *Annual Review of Microbiology*, 63, 541-546.
- Ma, Y., Wang, L., & Shao, Z. (2006). *Pseudomonas*, the dominant polycyclic aromatic hydrocarbon-degrading bacteria isolated from Antarctic soils and the role of large plasmids in horizontal gene transfer. *Environmental Microbiology*, 8(3), 455-465.
- Mac Cormack, W. P., & Fraile, E. R. (1997). Characterization of a hydrocarbon degrading psychrotrophic Antarctic bacterium. *Antarctic Science*, 9(2), 150-155.
- Macoustra, G. K., King, C. K., Wasley, J., Robinson, S. A., & Jolley, D. F. (2015). Impact of hydrocarbons from a diesel fuel on the germination and early growth of sub-Antarctic plants. *Environmental Science Processes and Impacts*, 17(7), 1238-1248.
- Malavenda, R., Rizzo, C., Michaud, L., Gerçe, B., Bruni, V., Syldatk, C., et al. (2015). Biosurfactant production by Arctic and Antarctic bacteria growing on hydrocarbons. *Polar Biology*, 38, 1565-1574.
- Marchal, R., Penet, S., Solano-Serena, F., & Vandecasteele, J. P. (2003). Gasoline and diesel oil biodegradation. *Oil and Gas Science and Technology*, 58(4), 441-448.
- Mardis, E. R. (2008). Next-generation DNA sequencing methods. *Annual Review of Genomics and Human Genetics*, 9, 387-402.
- Margesin, R., & Schinner, F. (1999). Biological decontamination of oil spills in cold environments. *Journal of Chemical Technology and Biotechnology*, 74, 381-389.
- Margesin, R., Gander, S., Zacke, G., Gounot, A. M., & Schinner, F. (2003). Hydrocarbon degradation and enzyme activities of cold-adapted bacteria and yeasts. *Extremophiles*, 7(6), 451-458.
- Marguiles, M., Egholm, M., Altman, W. E., Attiya, S., Bader, J. S., Bemben, L. A., Berka, J., et al. (2005). Genome sequencing in microfabricated high-density picolitre reactors. *Nature*, 437(7057), 376-380.
- Mariano, A. P., Tomasella, R. C., de Oliveira, L. M., Contiero, J., & de Angelis, D. F. (2008). Biodegradability of diesel and biodiesel blends. *African Journal of Biotechnology*, 7(9), 1323-1328.
- Marshall, B., Robleto, E. A., Wetzler, R., Kulle, P., Casaz, P., & Levy, S. B. (2001). The adnA transcriptional factor affects persistence and spread of *Pseudomonas fluorescens* under natural field conditions. *Applied Environmental Microbiology*, 67, 852-857.

- Martinez, Salette, & Hausinger, R. P. (2015). Catalytic mechanisms of Fe (II)- and 2-oxoglutarate-dependent oxygenases. *The Journal of Biological Chemistry*, 290(34), 20702-20711.
- Martinez-Alvarez, L. M., Ruberto, L. A. M., Lo Balbo, A., & Mac Cormack, W. P. (2017). Bioremediation of hydrocarbon-contaminated soils in cold regions: development of a pre-optimized biostimulation biopile-scale field assay in Antarctica. *Science of the Total Environment*, 90-91, 194-203. doi: 10.1016/j.scitotenv.2017.02.204.
- Masotti, A., & Preckel, T. (2006). Analysis of small RNAs with the Agilent 2100 Bioanalyzer. *Nature Methods/ Application Notes*.
- McDonald, R., & Knox, G. G. O. (2014). Cold region bioremediation of hydrocarbon contaminated soils: do we know enough?. *Environmental Science and Technology*, 48(17), 9980-9981.
- Meliani, A., & Bensoltane, A. (2014). Review of *Pseudomonas* attachment and biofilm formation in food industry. *Poultry, Fisheries and Wildlife Sciences*, 2(126), doi: 10.4172/2375-446X.1000126.
- Mengel, K., & Kirkby, E. A. (1987). Principles of Plant Nutrition. International Potash Institute, Worblaufen-Bern, Switzerland.
- Meyer, M., & Kircher, M. (2010). Illumina sequencing library preparation for highly multiplexed target capture and sequencing. *Cold Springs Harbor protocols*. doi: 10.1101/pdb.prot5448.
- Meyer, J-M., Stintzi, A., Coulanges, V., Shivaji, S., Voss, J. A., Taraz, K., & Budzikiewicz (1998). Siderotyping of fluorescent pseudomonads: characterization of pyoverdines of *Pseudomonas fluorescens* and *Pseudomonas putida* strains from Antarctica. *Microbiology*, 144, 3119-3126.
- Michaud, L., Lo Giudice, A., Saitta, M., De Domenico, M., & Bruni, V. (2004). The biodegradation efficiency on diesel oil by two psychrotrophic Antarctic marine bacteria during a two-long experiment. *Marine Pollution Bulletin*, 49, 405-409.
- Mille, G., Almallah, M., Bianchi, M., van Wambeke, F., & Bertrand, J. C. (1991). Effect of salinity on petroleum biodegradation, *Fresenius Journal of Analytical Chemistry*, 339, 788-791.
- Molavian, H. R., Kohandel, M., & Sivaloganathan, S. (2016). High concentrations of H₂O₂ make aerobic glycolysis energetically more favorable for cellular respiration. *Frontiers in Physiology*, 7, 362. doi: 10.3389/fphys.2016.00362.
- Montagnolli, R. N., Lopes, P. R. M., & Bidoia, E. D. (2015). Screening the toxicity and biodegradability of petroleum hydrocarbons by a rapid colorimetric method. *Archives of Environmental Contamination and Toxicology*, 68, 342–353.

- Morgan, P., & Watkinson, R. J. (1994). Biodegradation of components of petroleum. In Rathledge, C. (Ed), *Biochemistry of Microbial Degradation* (pp. 1-31). Dordrecht, Netherlands: Kluwer Academic Publishers.
- Morita, R. Y. (1975). Psychrophilic bacteria. *Bacteriological Reviews*, 37, 144–167.
- Muangchinda, C., Chavanich, S., Viyakara, V., Watanabe, K., Imura, S., Vangnai, A. S., et al. (2015). Abundance and diversity of functional genes involved in the degradation of aromatic hydrocarbons in Antarctic soils and sediments around Syowa Station. *Environmental Science and pollution Research*, 22, 4725-4735.
- Muhammad, S., Muller, T., & Jorgensen, R. G. (2008). Relationships between soil biological and other soil properties in saline and alkaline arable soils from the Pakistani Punjab. *Journal of Arid Environments*, 72, 448-457.
- Mukherjee, S., Huntemann, M., Ivanova, N., Kyrpides, N. C., & Pati, A. (2015). Large-scale contamination of microbial isolate genomes by Illumina PhiX control. *Standards in Genomic Sciences*, 10(18). doi: 10.1186/1944-3277-10-18.
- Mulder, H., Breure, A. M., Van Andel, J. G., Grotenhuis, J. T., & Rulkens, W. H. (1998). Influence of hydrodynamic conditions on naphthalene dissolution and subsequent biodegradation. *Biotechnology and Bioengineering*, 57(2), 145–154.
- Müller, T., Walter, B., Wirtz, A., & Burkovski, A. (2006). Ammonium toxicity in bacteria. *Current Microbiology*, 52(5), 400-406.
- Munoz, P. A., Marquez, S. L., Gonzalez-Nilo, F. D., Marquez-Miranda, V., & Blamey, J. M. (2017). Structure and application of antifreeze proteins from Antarctic bacteria. *Microbial Cell Factories*, 16(138), 1-13.
- Myers, R. H., & Montgomery, D. C. (1995). Response surface methodology: process and product optimization using designed experiments, John Wiley and Sons, New York.
- Nakamura, H., Kobayashi, S., Hirata, Y., Suzuki, K., Mogi, Y., & Karube, I. (2007). A spectrophotometric biochemical oxygen demand determination method using 2,6-dichlorophenolindophenol as the redox color indicator and the eukaryote *Saccharomyces cerevisiae*. *Analytical Biochemistry*, 369, 168-174.
- Nakamura, F. M., Germano, M. G., & Tsai, S. M. (2014). Capacity of aromatic compound degradation by bacteria from Amazon Dark Earth. *Diversity*, 6, 339–353. doi: 10.3390/d6020339.
- Naseby, D. C., Way, J. A., Bainton, N. J., & Lynch, J. M. (2001). Biocontrol of *Phytophthora* in the pea rhizosphere by antifungal metabolite producing and non-producing *Pseudomonas* strains. *Journal of Applied Microbiology*, 90(3), 421-429.

- Nie, Y., Chi, C. Q., Fang, H., Liang, J. L., Lu, S. I., Lai, G. L., et al. (2014). Diverse alkane hydroxylase genes in microorganisms and environments. *Scientific Reports*, 4, 4968.
- Nkem, J. N., Virginia, R. A., Barrett, J. E., Wall, D. H., & Li, G. (2006). Salt tolerance and survival thresholds for two species of Antarctic soil nematodes. *Polar Biology*, 29(8), 643–651. <https://doi.org/10.1007/s00300-005-0101-6>
- NRC (National Research Council) (2003). Oil in the sea III: inputs, fates and effects. Washington: The National Academic Press.
- Nydahl, A. C., King, C. K., Wasley, J., Jolley, D. F., & Robinson, S. A. (2015). Toxicity of fuel-contaminated soil to Antarctic moss and terrestrial algae. *Environmental Toxicology and Chemistry*, 34(9), 2004–2012.
- Nyren, P. (1987). Enzymatic method for continuous monitoring of DNA polymerase activity. *Analytical Biochemistry*, 167(2), 235-238.
- Nyren, P., & Lundin, A. (1985). Enzymatic method for continuous monitoring of inorganic pyrophosphate synthesis. *Analytical Biochemistry*, 151(2), 504-509.
- Obidike, I. R., Maduabuchi, I. U., & Olumuyiwa, S. S. O. (2007). Testicular morphology and cauda epididymal sperm reserves of male rats exposed to Nigerian Qua Iboe Brent crude oil. *Oil and Chemical Pollution*, 3, 117–129.
- Oboh, B. O., Adeyinka, Y., Awonuga, S., & Akinola, M. O. (2007). Impact of soil types and petroleum effluents on the earthworm *Eudrilus eugeniae*. *Journal of Environmental Biology*, 28(2), 209–212.
- OEHHA (Office of Environmental Health Hazard Assessment) (2001). Health effects of diesel exhaust. Sacramento, CA. California Environmental Protection Agency.
- Okpokwasili, G. C., Somerville, C. C., Sullivan, M., Grimes, D. J., & Colwell, R. R. (1986). Plasmid-mediated degradation of hydrocarbons by estuarine bacteria. *Research Journal of Environmental Toxicology*, 3(1), 9–23.
- Ondov, B. D., Bergman, N. H., & Phillippy, A. M. (2011). Interactive metagenomic visualization in a Web browser. *BMC Bioinformatics*, 12(385). doi: 10.1186/1471-2105-12-385.
- Oudot, J., Merlin, F. X., & Oinvidic, P. (1998). Weathering rates of oil components in a bioremediation experiment in estuarine sediments. *Marine Environmental Research*, 45(2), 113-125.
- Overbeek, R., Olson, R., Pusch, G. D., Olsen, G. J., Davis, J. J., Disz, T., Edwards, R. A., et al. (2014). The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). *Nucleic Acid Research*, 42, 206-214. <https://doi.org/10.1093/nar/gkt1226>.

- Pacheco, G. J., Ciapina, E. M., Gomes, E. B., & Junior, N. P. (2010). Biosurfactant production by *Rhodococcus erythropolis* and its application to oil removal. *Brazilian Journal of Microbiology*, 41(3), 685-693. doi: 10.1590/S1517-83822010000300019.
- Pacwa-Plociniczak, M., Plaza, G. A., Poliwoda, A., & Piotrowska-Seget, Z. (2014). Characterization of hydrocarbon-degrading and biosurfactant-producing *Pseudomonas* sp. P-1 strain as a potential tool for bioremediation of petroleum-contaminated soil. *Environmental Science and Pollution Research*, 21, 9385–9395.
- Paixao, J. F., Nascimento, I. A., Pereira, S. A., Leite, M. B. L., Carvalho, G. C., Silveira Jr, J. S. C., et al. (2007). Estimating the gasoline component and formulations toxicity to microalgae (*Tetramis chuii*) and oyster (*Crassostrea rhizophorae*) embryos: an approach to minimize environmental pollution risk. *Environmental Research*, 103(3), 365-374.
- Pandey, G., & Jain, R. K. (2002). Bacterial chemotaxis toward environmental pollutants: role in bioremediation. *Applied and Environmental Microbiology*, 68(12), 5789-5795.
- Panke, S., Sanchez-Romero, J. M., & de Lorenzo, V. (1998). Engineering of quasi-natural *Pseudomonas putida* strains for toluene metabolism through an ortho-cleavage degradation pathway. *Applied and Environmental Microbiology*, 64(2), 748–751.
- Parales, R. E., & Harwood, C. S. (2002). Bacterial chemotaxis to pollutants and plant-derived aromatic molecules. *Current Opinion in Microbiology*, 5(3), 266-273.
- Parales, R. E., Parales, J. V., Pelletier, D. A., & Ditty, J. L. (2008). Diversity of microbial toluene degradation pathways. *Advances in Applied Microbiology*, 64, 1-73.
- Perelo, L. W. (2010). Review: *in situ* and bioremediation of organic pollutants in aquatic sediments. *Journal of Hazardous Materials*, 177, 81–89.
- Peressutti, S. R., Alvarez, A. M., & Pucci, O. H. (2003). Dynamics of hydrocarbon-degrading bacteriocenosis of an experiment oil pollution in Patagonian soil. *International Biodeterioration and Biodegradation*, 52, 21–30.
- Perez-de-Mora, A., Engel, M., & Schloter, M. (2011). Abundance and diversity of *n*-alkane degrading bacteria in a forest soil co-contaminated with hydrocarbons and metals: a molecular study on alkB homologous gene. *Microbial Ecology*, 62(4), 959–972.
- Pérez-Pantoja, D., Gonzalez, B., & Pieper, D. H. (2010). Aerobic degradation of aromatic hydrocarbons. In Timmis, K. N. (Ed), *Handbook of Hydrocarbon and Lipid Microbiology*. doi: 10.1007/978-3-540-77587-4_60, Springer-Verlag Berlin Heidelberg.

- Peterson, D. R. (1994). Calculating the aquatic toxicity of hydrocarbon mixtures. *Chemosphere*, 29(12), 2493–2506.
- Pieper, D. H. (2005). Aerobic degradation of polychlorinated biphenyls. *Applied Microbiology and Biotechnology*, 67(2), 170–191.
- Powell, S. M., Ferguson, S. H., Snape, I., & Siciliano, S. D. (2006). Fertilization stimulates anaerobic fuel degradation of Antarctic soils by denitrifying microorganisms. *Environmental Science and Technology*, 40(15), 2011–2017.
- Powell, S. M., Harvey, P. M., Stark, J. S., Snape, I., & Riddle, M. J. (2007) Biodegradation of petroleum products in experimental plots in Antarctic marine sediments is location dependent. *Marine Pollution Bulletin*, 54(4), 434-440.
- Peng, X., Taki, H., Komukai, S., Sekine, M., Kanoh, K., Kasai, H., Choi, S-K., et al. (2006). Characterization of four *Rhodococcus* alcohol dehydrogenase genes responsible for the oxidation of aromatic alcohols. *Applied Microbiology and Biotechnology*, 71, 824-832.
- Perry, J. J. (2007). Microbial metabolism of cyclic alkanes. In Atlas, R. M. *Petroleum Microbiology*. New York, Macmillan Publishing Co., 61-97.
- Plackett, R. L., & Burman, J. P. (1946). The Design of Optimum Multifactorial Experiments. *Biometrika*, 33(4), 305–325. <https://doi.org/10.2307/2332195>
- Poland, J. S., Riddle, M. J., & Zeeb, B. A. (2003). Contaminants in the Arctic and Antarctic: a comparison of sources, impacts, and remediation options. *Polar Record*, 39, 369-384.
- Prosser, J. I., & Embley, T. M. (2002). Cultivation-based and molecular approaches to characterisation of terrestrial and aquatic nitrifiers. *Antonie Van Leeuwenhoek*, 81(1-4), 165-179.
- Provint, T., & Pitt, J. L. (2001). Managing soil salinity. Texas A and M University, Texas: Texas Agricultural Extension Service, pp 5.
- Rabinowitz, J. D., Vacchino, J. F., Beeson, C., & McConnell, H. M. (1998). Potentiometric measurements of intracellular redox activity. *Journal of the American Chemical Society*, 120, 2464-2473.
- Rabus, R., Boll, M., Heider, J., Meckenstock, R. U., Buckel, W., Einsle, O., Ermler, U., et al (2016). Anaerobic microbial degradation of hydrocarbons: from enzymatic reactions to the environment. *Journal of Molecular Microbiology and Biotechnology*, 26(1-3), 5–28. doi: 10.1159/000443997.
- Rainey, F. A., Burghardt, J., Kroppenstedt, R. M., Klatte, S., & Stackerbrandt, E. (1995). Phylogenetic analysis of the genera *Rhodococcus* and *Nocardia* and evidence for the evolutionary origin of the genus *Nocardia* from within the radiation of *Rhodococcus* species. *Microbiology*, 141, 523-528.

- Rainwater, F. H., & Thatcher, L. L. (1960). *Methods for Collection and Analysis of Water Samples* (Water Supply Paper No. 1454) (p. 301). U.S.: U.S. Government Printing Office.
- Rajendhran, J., & Gunasekaran, P. (2011). Microbial phylogeny and diversity: small subunit ribosomal RNA sequence analysis and beyond. *Microbiological Research*, 166(2), 99-110.
- Raymond, T., King, C. K., Raymond, B., Stark, J. S., & Snape, I. (2017). Chapter 14 - Oil Pollution in Antarctica. In M. Fingas (Ed.), *Oil Spill Science and Technology (Second Edition)* (pp. 759–803). Boston: Gulf Professional Publishing.
- Rayner, J. L., Snape, I., Walworth, J. L., Harvey, P. M., & Ferguson, S. H. (2007). Petroleum-hydrocarbon contamination and remediation by microbioventing at sub-Antarctic Macquarie Island. *Cold Regions Science and Technology*, 48, 139-153.
- Reddy, G. S., Matsumoto, G. I., Schumann, P., Stackerbrandt, E., & Shivaji, S. (2004). Psychrophilic pseudomonads from Antarctica: *Pseudomonas antarctica* sp. nov., *Pseudomonas meridiana* sp. nov. and *Pseudomonas proteolytica* sp. nov. *International Journal of Systematic and Evolutionary Microbiology*, 54(Pt 3), 713-719.
- Richard, J. Y., & Vogel, T. M. (1999) Characterization of a soil bacterial consortium capable of degrading diesel fuel. *International Biodeterioration and Biodegradation*, 44(2-3), 93-100.
- Richardson, E. L., King, C. K., & Powell, S. M. (2014) The use of microbial gene abundance in the development of fuel remediation guidelines in polar soils. *Integrated Environmental Assessment and Management*, 11(2), 235-241.
- Rhykerd, R. I., Weaver, R. W., & McInnes, K. J. (1995). Influence of salinity on bioremediation of oil in soil. *Environmental Pollution*, 90, 127-130.
- Rietz, D. N., & Haynes, R. J. (2003). effects of irrigation-induced salinity and sodicity on soil microbial activity. *Soil Biology and Biochemistry*, 35(6), 845-854.
- Rodriguez-Fernandez, J., Pereiro, R., & Sanz-Mendel, A. (2002). Optical fibre sensor for hydrogen sulphide monitoring in mouth air. *Analytica Chimica Acta*, 471, 13-23.
- Rodriguez-Trigo, G., Zock, J., Pozo-Rodriguez, F., Gomez, F. P., Monyarch, G., Bouso, L., et al. (2010). Health changes in fishermen 2 years after clean-up of the Prestige oil spill. *Annals of Internal Medicine*, 153(8), 489-498.
- Rojo, F. (2009). Degradation of alkanes by bacteria. *Environmental Microbiology*, 11(10), 2477–2490.

- Ruberto, L., Vazquez, S., Lo Balbo, A., & Mac Cormack, W. P. (2005). Psychrotolerant hydrocarbon-degrading *Rhodococcus* strains isolated from polluted Antarctic soils. *Antarctic Science*, 17(1), 47-56.
- Ruberto, L., Vazquez, S. C., & Mac Cormack, W. P. (2003). Effectiveness of the natural bacterial flora, biostimulation and bioaugmentation on the bioremediation of the hydrocarbon contaminated Antarctic soil. *International Biodegradation and Biodegradation*, 52(2), 115-125.
- Ronaghi, M., Karamohamed, S., Pettersson, B., Uhlen, M., & Nyren, P. (1996). Real-time DNA sequencing using detection of pyrophosphate release. *Analytical Biochemistry*, 242(1), 84-89.
- Ronaghi, M., Uhlen, M., & Nyren, P. (1998). A sequencing method based on real-time pyrophosphate. *Science*, 281(5375), 363-365.
- Saito, A., Iwabuchi, T., & Harayama, S. (1999). Characterization of genes for enzymes involved in the phenanthrene degradation in *Nocardioides* sp. KP7, *Chemosphere*, 38, 1331-1337.
- Saitou, N., & Nei, M. (1987). The neighbour-joining method: a new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution*, 4(4), 406-425.
- Sanger, F., Nicklen, S., & Coulson, A. R. (1977). DNA sequencing with chain-terminating inhibitors, *Proceedings of the National Academy Sciences USA*, 74, 5463-5467.
- Santa Anna, L. M., Sebastian, G. V., Menezes, E. P., Alves, T. L. M., Santos, A. S., Junior, N. P., et al. (2002) Production of biosurfactants from *Pseudomonas aeruginosa* PA1 isolated in oil environments. *Brazilian Journal of Chemical Engineering*, 19(2), 159-166.
- Saul, D. J., Aislabie, J. M., Brown, C. E., Harris, L., & Foght, J. M. (2005). Hydrocarbon contamination changes the bacterial diversity of soil from around Scott Base, Antarctica. *FEMS Microbiology Ecology*, 53, 141-155.
- SCAR (Scientific Committee on Antarctic Research) (1993). Protocol on environmental protection to the Antarctic Treaty. *Polar Record*, 29, 256-275.
- Schweitzer, C., & Scaiano, J. C. (2003). Selective binding and local photophysics of the fluorescent cyanine dye PicoGreen in double-stranded and single-stranded DNA. *Physical Chemistry Chemical Physics*, 5(21), 4911-4917.
- Sekine, M., Tanikawa, S. S., Omata, S., Saito, M., Fujisawa, T., Tsukatani, N., et al. (2006). Sequence analysis of three plasmids harboured in *Rhodococcus erythropolis* strain PR4. *Environmental Microbiology*, 8(2), 334-346.
- Sierra-Garcia, I. N., & de Oliveira, V. M. (2013). Microbial hydrocarbon degradation: efforts to understand biodegradation in petroleum reservoirs. *Biodegradation - Engineering and Technology*, doi: 10.5772/55920.

- Seo, J-S., Keum, Y-S., & Li, Q. X. (2009). Bacterial degradation of aromatic compounds. *International Journal of Environmental Research and Public Health*, 6(1), 278-309.
- Silby, M. W., & levy, S. B. (2004). Use of in vivo expression technology to identify genes important in growth and survival of *Pseudomonas fluorescens* Pf0-1 in soil: discovery of expressed sequences with novel genetic organization. *Journal of Bacteriology*, 186(21), 7411-7419.
- Silby, M. W., & Levy, S. B. (2008). Overlapping protein-encoding genes in *Pseudomonas fluorescens* Pf0-1. *PLoS Genetics*, 4(6), e1000094.
- Silby, M. W., Nicoll, J. S., & Levy, S. B. (2009). Requirement of polyphosphate by *Pseudomonas fluorescens* Pf0-1 for competitive fitness and heat tolerance in laboratory media and sterile soil. *Applied and Environmental Microbiology*, 75(12), 3872-3881.
- Singh, P., DeMarini, D. M., Dick, C. A. J., Tabor, D. G., Ryan, J. V., Linak, W. P., et al. (2004). Sample characterisation of automobile and forklift diesel exhaust particles and comparative pulmonary toxicity in mice. *Environmental Health Perspectives*, 112(8), 820-825.
- Singh, A., & Ward, O. P. (2004). Biotechnology and bioremediation- an overview. In: Singh, A., Ward, O. P. (ed.) Biodegradation and Bioremediation. *Soil Biology*, vol. 2. Springer-Verlag, Berlin, Heidelberg.
- Shimizu, K. (2013). Metabolic Regulation of a Bacterial Cell System with Emphasis on Escherichia coli Metabolism. *International Scholarly Research Notices*. Retrieved from <https://www.hindawi.com/journals/isrn/2013/645983/>
- Shukor, M. Y., Hassan, N. A., Jusoh, A. Z., Perumal, N., Shamaan, N. A., Mac Cormack, W. P., et al. (2009). Isolation and characterisation of a *Pseudomonas* diesel-degrading strain from Antarctica. *Journal of Environmental Biology*, 30(1), 1-6.
- Siqueira, J. F., Fouad, A. F., & Rôcas, I. N. (2012). Pyrosequencing as a tool for better understanding of human microbiomes. *Journal of Oral Microbiology*, 4. <https://doi.org/10.3402/jom.v4i0.10743>
- Snape, I., Morris, C. E., & Cole, C. M. (2001). The use of permeable reactive barriers to control contaminants dispersal during site remediation in Antarctica. *Cold Regions Science and Technology*, 32, 157-174.
- Snape, I., Acomb, L., Barnes, D. I., Bainbridge, S., Eno, R., Filler, D. M., Plato, N., et al. (2008). Contamination, regulation and remediation: an introduction to bioremediation of petroleum hydrocarbons in cold regions. In Filler, D. M., Snape, I., & Barnes, D. L. *Bioremediation of petroleum hydrocarbons in cold regions*, Cambridge University Press.
- Staden, R. (1980). A new computer method for the storage and manipulation of DNA gel reading data. *Nucleic Acids Research*, 8(16), 3673-3694.

- Stallwood, B., Shears, J., Williams, P. A., & Hughes, K. A. (2005). Low temperature bioremediation of oil-contaminated soil using biostimulation and bioaugmentation with a *Pseudomonas* sp. from maritime Antarctica. *Journal of Applied Microbiology*, 99(4), 794-802.
- Stancu, M. M. (2015). Response of *Rhodococcus erythropolis* strain IBB_{pol} to toxic organic solvents. *Brazilian Journal of Microbiology*, 46(4), 1009-1018.
- Stevens, V., Thijs, S., McAmmond, B., Langill, T., Van Hamme, J., Weyens, N., & Vangronsveld, J. (2017). Draft genome sequence of *Rhodococcus erythropolis* VSD3, a diesel fuel-degrading and plant growth-promoting bacterium isolated from *Hedera helix* leaves. *Genome Announcements*, 5(8). doi: 10.1128/genomeA.01680-16.
- Strong, M. J., Xu, G., Morici, L., Bon-Durant, S. S., Baddoo, M., Lin, Z., Fewell, C., Taylor, C. M., & Flemington, E. K. (2014). Microbial contamination in next generation sequencing: implications for sequence-based analysis of clinical samples. *PLoS Pathogen*, 10(11): e1004437. doi: 10.1371/journal.ppat.1004437.
- Suyama, A., Iwakiri, R., Kimura, N., Nishi, A., Nakamura, K., & Furukawa, K. (1996). Engineering hybrid pseudomonads capable of utilizing a wide range of aromatic hydrocarbons and of efficient degradation of trichloroethylene. *Journal of Bacteriology*, 178(14), 4039-4046.
- Swerdlow, H., & Gesteland, R. (1990). Capillary gel electrophoresis for rapid, high resolution DNA sequencing. *Nucleic Acids Research*, 18(6), 1415-1419.
- Taketani, R. G., Zucchi, T. D., de Melo, I. S., & Mendes, R. (2013). Whole-genome shotgun sequencing of *Rhodococcus erythropolis* strain P27, a highly radiation-resistant actinomycete from Antarctica. *Genome Announcement*, 1(5), e00763-13.
- Takahara, H., Ogihara, J., Yoshida, T., Okuda, S., Nakajima, M., Iwabuchi, N., & Sunain, M. (2014). Enhanced translocation and growth of *Rhodococcus erythropolis* PR4 in the alkane phase of aqueous-alkane two phase cultures were mediated by GroEL2 overexpression. *Microbes and Environment*, 29(4), 346–352.
- Tanco, M., Viles, E., & Pozueta, L. (2009). Comparing Different Approaches for Design of Experiments (DoE). In *Advances in Electrical Engineering and Computational Science* (pp. 611–621). Springer, Dordrecht.
- Tang, J., Wang, M., Wang, F., Sun, Q., & Zhou, Q. (2011). Eco-toxicity of petroleum hydrocarbon contaminated soil. *Journal of Environmental Sciences*, 23(5), 845-851.
- Tawfik, D. S., & Griffiths, A. D. (1998). Man-made cell-like compartments for molecular evolution. *Nature Biotechnology*, 16(7), 652-656.

- Teare, J. M., Islam, R., Flanagan, R., Gallagher, S., Davies, M. G., & Grabeau, C. (1997). Measurement of nucleic acid concentrations using the DyNA QuantTM and the GeneQuantTM. *Biotechniques*, 22, 1170-1174.
- Tomei, M. C., & Daugulis, A. J. (2013). *Ex situ* bioremediation of contaminated soils: an overview of conventional and innovative technologies. *Critical Reviews in Environmental Science and Technology*, 43, 2107-2139.
- Tourova, T. P., Sokolova, D. S., Semenova, E. M., Shumkova, E. S., Korhunova, A. V., Babich, T. L., Poltaraus, A. B., & Nazina, N. (2016). Detection of *n*-alkane biodegradation genes *alkB* and *ladA* in thermophilic hydrocarbon-oxidizing bacteria of the genera *Aeribacillus* and *Geobacillus*. *Microbiology*, 85(6), 693-707.
- Tripathi, S., Kumari, S., Chakraborty, A., Gupta, A., Chakrabarti, K., & Bandyapadhyay, B. K. (2006). Microbial biomass and its activities in salt-affected coastal soils. *Biology and Fertility of Soils*, 42, 273-277.
- Turcatti, G., Romieu, A., Fedurco, M., & Tairi, A. P. (2008). A new class of cleavable fluorescent nucleotides: synthesis and optimization as reversible terminators for DNA sequencing by synthesis. *Nucleic Acids Research*, 36(4), e25. doi: 10.1093/nar/gkn021.
- Tyagi, M., da Fonseca, M. M. R., & de Carvalho, C. C. C. R. (2011). Bioaugmentation and biostimulation strategies to improve the effectiveness of bioremediation processes. *Biodegradation*, 22(2), 231-241.
- Ulrich, A. C., Guigard, S. E., Foght, J. M., Semple, K. M., Pooley, K., Armstrong, J. E., & Biggar, K. W. (2006). Effect of salt on aerobic biodegradation of petroleum hydrocarbons in contaminated groundwater. *Biodegradation*, 20, 27-38.
- Urai, M., Yoshizaki, H., Anzai, H., Ogihara, J., Iwabuchi, N., Harayama, S., Sunairi, M., & Nakajima, M. (2007). Structural analysis of an acidic, fatty acid ester-bonded extracellular polysaccharides produced by pristane-assimilating marine bacterium, *Rhodococcus erythropolis* PR4. *Carbohydrate Research*, 342, 933-942.
- van Beilen, J. B., Li, Z., Duetz, W. A., Smits, T. H. M., & Witholt, B. (2003). Diversity of alkane hydroxylase systems in the environment. *Oil and Gas Science and Technology*, 58, 427-440.
- van Beilen, J. B., & Funhoff, E. G. (2007). Alkane hydroxylases involved in microbial alkane degradation. *Applied Microbiology and Biotechnology*, 74(1), 13-21.
- Van Gestel, K., Mergaert, J., Swings, J., Coosemans, J., & Ryckeboer, J. (2003). Bioremediation of diesel oil-contaminated soil by composting with biowaste. *Environmental Pollution*, 125, 361-368. doi: 10.1016/S0269-7491(03)00109-X.
- Van Hamme, J. D., Singh, A., & Ward, O. P. (2003). Recent advances in petroleum microbiology. *Microbiology and Molecular Biology Review*, 67(4), 503-549.

- Van Sluys, M. A., Monteiro-Vitorello, C. B., Camargo, L. E., Menck, C. F., Da Silva, A. C., Ferro, J. A., Oliveira, M. C., Setubal, J. C., Kitajima, J. P., & Simpson, A. J. (2002). Comparative genomic analysis of plant-associated bacteria. *Annual Review of Phytopathology*, 40, 169-189.
- Varivarn, K., Champa, L. A., Silby, M. W., & Robleto, E. A. (2013). Colonization strategies of *Pseudomonas fluorescens* Pf0-1: activation of soil-specific genes important for diverse and specific environments. *BMC Microbiology*, 13(92). doi: 10.1186/1471-2180-13-92.
- Vasudevan, N., Bharathi, S., & Arulazhagan, P. (2007). Role of plasmid in the degradation of petroleum hydrocarbons by *Pseudomonas fluorescens* NS1. *Journal of Environmental Science and Health*, 42(8), 1141–1146.
- Vazquez, S., Nogales, B., Ruberto, L., Mestre, C., Christie-Oleza, J., Ferrero, M., Bosch, R., & Mac Cormack, W. P. (2013). Characterization of bacterial consortia from diesel-contaminated Antarctic soils: towards the design of tailored formulas for bioaugmentation. *International Biodeterioration and Biodegradation*, 77, 22–30.
- Venkatesh, S., & Dayananda, C. (2008). Properties, potentials, and prospects of antifreeze proteins. *Critical Reviews in Biotechnology*, 28, 57-82.
- Vilchez-Vargas, R., Junca, H., & Pieper, D. H. (2010). Metabolic networks, microbial ecology and 'omics' technologies: towards understanding *in situ* biodegradation processes. *Environmental Microbiology*, 12(12), 3089-3104.
- Voelkerding, K. V., Dames, S., & Durtschi, J. D. (2009). Next-generation-sequencing for clinical diagnostic-principles and application to targeted resequencing for hypertrophic cardiomyopathy. *The Journal of Molecular Diagnostics*, 12(5), 539–551.
- von Wirèn, N., Lauter, F-R., Ninnemann, O., Gillissen, B., Walch-Liu, P., Engels, C., Jost, W., & Frommer, W. B. (2000). Differential regulation of three functional ammonium transporter genes by nitrogen in root hairs and by light in leaves of tomato. *The Plant Journal*, 21(2), 167-175.
- von Wirèn, N., & Merrick, M. (2004). Regulation and function of ammonium carriers in bacteria, fungi, and plants. In: Molecular mechanisms controlling transmembrane transport. *Current Genetics*, vol 9. Springer, Berlin, Heidelberg.
- Walworth, J., Harvey, P., & Snape, I. (2013) Low temperature soil petroleum hydrocarbon degradation at various oxygen levels. *Cold Regions Science and Technology*, 96, 117-121.
- Walworth, J., Pond, A., Snape, I., Rayner, J., Ferguson, S., & Harvey, P. (2007) Nitrogen requirements for maximizing petroleum bioremediation in a sub-Antarctic soil. *Cold Regions Science and Technology*, 48, 84-91.

- Wang, X., & Bartha, R. (1990). Effects of bioremediation on residues, activity and toxicity in soil contaminated by fuel soils. *Soil Biology and Biochemistry*, 22(4), 501-505.
- Wang, W., & Shao, Z. (2013). Enzymes and genes involved in aerobic alkane degradation. *Frontiers in Microbiology*, 4(116).
- Warhurst, A. M., & Fewson, C. A. (1994). Biotransformations catalysed by the genus *Rhodococcus*. *Critical Reviews in Biotechnology*, 14(1).
- Ward, D. M., & Brock, T. D. (1978). Hydrocarbon biodegradation in hypersaline environments. *Applied Environmental Microbiology*, 35(2), 353-359.
- Webster, J., Webster, K., Nelson, P., & Waterhouse, E. (2003). The behaviour of residual contaminants at a former station site, Antarctica. *Environmental Pollution*, 123, 163–179.
- Wentzel, A., Ellingsen, T. E., Kotlar, H. K., Zotchev, S. B., & Throne-Holst, M. (2007). Bacterial metabolism of long-chain *n*-alkanes. *Applied Microbiology and Biotechnology*, 76, 1209-1221.
- White, D. A., Hird, L. C., & Ali, S. T. (2013). Production and characterization of a trehalolipid biosurfactant produced by the novel marine bacterium *Rhodococcus* sp., strain PML026. *Journal of Applied Microbiology*, 115(3), 744-755.
- Whyte, L. G., Slagman, S. J., Pietrantonio, F., Bourbonniere, L., Koval, S. F., Lawrence, J. R., Inniss, W. E., & Greer, C. W. (1999). Physiological adaptations involved in alkane assimilation at a low temperature by *Rhodococcus* strain Q15. *Applied and Environmental Microbiology*, 65(7), 2961-2968.
- Whyte, L. G., Smits, T. H., Labbè, D., Witholt, B., Greer, C. W., & Van Beilen, J. B. (2002) Gene cloning and characterisation of multiple alkane hydroxylase systems in *Rhodococcus* strain Q15 and NRRL B-16531. *Applied and Environmental Microbiology*, 68(12), 5933-5942
- Woese, C. R., & Fox, G. E. (1977). Phylogenetic structure of prokaryotic domain: the primary kingdom. *Proceedings of the National Academy of Sciences of the USA*, 74(11), 5088-5090.
- Woolfenden, E. N. M., Hince, G., Powell, S. M., Stark, S. C., Snape, I., Stark, J. S., & George, S. C. (2011). The rate of removal and the compositional changes of diesel in Antarctic marine sediment. *Science of Total Environment*, 410-411, 205-216.
- Wu, L., Wen, C., Qin, Y., Yin, H., Tu, Q., Van Nostrand, J. D., Yuan, T., et al. (2015). Phasing amplicon sequencing on Illumina MiSeq for robust environmental microbiology community analysis. *BMC Microbiology*, 15(25).

- Xu, X., & Zhu, X. (2004). Treatment of refractory oily wastewater by electro-coagulation process. *Chemosphere*, 56, 889-894.
- Yari, A., & Kargosha, K. (2006). Simple photometric determination of free cyanide ion in aqueous solution with 2,6-dichlorophenolindophenol. *Central European Journal of Chemistry*, 4(2), 329-337.
- Yang, S. Z., Jin, H. J., Wei, Z., He, R. X., Ji, Y. J., Li, X. M., & Yu, S. P. (2009). Bioremediation of oil spills in cold environments: a review. *Pedosphere*, 19(3), 371-381.
- Yeung, C. W., Van Stempoort, D. R., Spoelstra, J., Bickerton, G., Voralek, J., & Greer, C. W. (2013). Bacterial community evidence for anaerobic degradation of petroleum hydrocarbons in cold climate groundwater. *Cold Regions Science and Technology*, 86, 55-68.
- Yoshida, N., Hoashi, J., Morita, T., McNiven, S. J., Nakamura, H., & Karube, I. (2001). Improvement of a mediator-type biochemical oxygen demand sensor for on-site measurement. *Journal of Biotechnology*, 88, 269-275.
- Young, E., Stewart, F., & Dimalanta, E. (2017). The quantitation question: how does accurate library quantitation influence sequencing? *New England BioLabs*, Feature article.
- Zanaroli, G., Di Toro, S., Todaro, D., Varese, G. C., Bertolotto, A., & Fava, F. (2010). Characterization of two diesel fuel degrading microbial consortia enriched from a non acclimated, complex source of microorganisms. *Microbial Cell Factories*, 9(10). doi: 10.1186/1475-2759-9-10.
- Zerbino, D. R. (2010). Using the velvet *de novo* assembler for short-read sequencing technologies. *Current Protocols in Bioinformatics*, 11(5), 1-13.
- Zerbino, D. R., & Birney, E. (2008). Velvet: algorithm for the *de novo* short read assembly using de Bruijn graphs. *Genome Research*, 18(5), 821-829.
- Zock, J. P., Rodriguez-Trigo, G., Pozo-Rodriguez, F., Barbera, J. A., Bouso, L., Torralba, Y., et al. (2007). Prolonged respiratory symptoms in clean-up workers of the Prestige oil spill. *American Journal of Respiratory and Critical Care Medicine*, 176(6), 610-616.
- Zock, J. P., Rodriguez-Trigo, G., Rodriguez-Rodriguez, E., Pozo-Rodriguez, F., Gomez, F. P., Fuster, C., et al. (2009). Long-term health effects of the Prestige oil spill (Galicia, Spain). *Epidemiology*, 20(6), 242-243.
- Zock, J. P., Rodriguez-Trigo, G., Rodriguez-Rodriguez, E., Espinosa, A., Pozo-Rodriguez, F., Gomez, F. P., et al. (2012). Persistent respiratory symptoms in clean-up workers 5 years after the Prestige oil spill. *Occupational and Environmental Medicine*, 69(7), 508-513.